Detection of ZO-1 in Formalin-Fixed, Paraffin-Embedded Rat Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
Protease
Normal Rabbit IgG – Affinity Purified
DAB Chromogen
Hematoxylin

Blocking Serum: Normal Donkey Serum
Jackson Immunoresearch Laboratories, Inc.
West Grove, PA 19390
www.jacksonimmuno.com
1-800-367-5296
Catalog # 017-000-001

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rabbit Polyclonal Anti-ZO-1 Antibody Invitrogen Carlsband, CA 92008 www.invitrogen.com

www.invitrogen.com 1-800-955-6288 Catalog # 61-7300

Secondary Antibody: Biotin-Conjugated Donkey Anti-Rabbit IgG

Jackson Immunoresearch Laboratories, Inc.

West Grove, PA 19390 www.jacksonimmuno.com 1-800-367-5296 Catalog # 711-065-152

Label Complex: R.T.U. Vectastain Elite ABC Reagent

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # PK-7100

Staining Procedure

Positive Control Tissue: Kidney (capillary loops in glomeruli)

Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Solution	Repetitions	Time
Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.
- 4. <u>Proteolytic-Induced Epitope Retrieval Using Protease</u>
 Incubate the slides in a 0.01% protease solution (made in distilled water) for 5 minutes at 37°C.
 Rinse the slide in distilled water for 1 minute to stop the enzymatic reaction.

 5. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each time.

6. Block with 10%	normal donkey serum for 20 minutes at room temperature
Lot #	Date Reconstituted
DO NOT RINSE	SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK.

7. Avidin / Biotin Blocking Kit

Lot #_____ Exp. Date____ New Kit: yes / no
Apply avidin block for 15 minutes at room temperature.

Quick rinse in 1X wash buffer.

Apply biotin block for 15 minutes at room temperature.

DO NOT RINSE SECTIONS WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BLOCK.

8. Apply 1	rimary antibody at a 1:25 dilution. Incubate for 1 hour at room temperature.
Lot #	Exp. Date

For negative control slides, dilute normal rabbit IgG so that it's IgG protein concentration matches that of the primary antibody (if necessary). Then make a 1:25 dilution. If the concentrations can't be matched using this method, the dilution for the negative reagent may need to be adjusted. Apply the negative and incubate for 1 hour at room temperature.

Lot #	Exp. Date	

9. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.

10. Apply the donkey anti-rabbit secondary antibody at a 1:1000 dilution. Incubate for 30 minutes at room temperature.
Lot # Date Reconstituted
11. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.
12. Apply the Vectastain R.T.U. Elite Label and incubate for 30 minutes at room temperature. Exp. Date New Kit: yes / no
13. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each time.
14. Apply the DAB chromogen. Incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate)
Lot # Exp. Date New Kit: yes / no
15. Rinse the slides in tap water 3 minutes.
16. Counterstain with hematoxylin for 20 seconds.
17. Rinse the slides in tap water until water is clear.
18. Gently agitate slides in 1X wash buffer until the tissues turn blue.

Solutions	Repetitions	Time
95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

19. Dehydrate through the following solutions:

20. Coverslip

Updated 10/15/07