Detection of Cytokeratin 20 in Formalin-Fixed, Paraffin-Embedded Rat Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
1X Citrate Buffer
Normal Rabbit IgG – Affinity Purified
DAB Chromogen
Hematoxylin

Staining Kit: Vectastain Elite ABC Kit (Rabbit IgG)

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # PK-6101

Note: This kit contains all reagents necessary to make the blocking reagent, secondary antibody and label complex.

Avidin / Biotin Blocking Kit Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rabbit Polyclonal Anti-Cytokeratin 20 Antibody

Abcam, Inc Cambridge, MA 02139 www.abcam.com 1-888-772-2226 Catalog # ab53120

Staining Procedure

Positive Control Tissue: Gastrointestinal tract Stain Localization: Cytoplasmic and cell membrane

1. Deparaffinize and hydrate slides through the following solutions:

| Solution | Repetitions | Time |
|----------------|-------------|-----------|
| Xylene | 2 times | 5 minutes |
| 100% Ethanol | 2 times | 3 minutes |
| 95% Ethanol | 2 times | 3 minutes |
| 1X Wash Buffer | 2 times | 5 minutes |

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.

| 1. | Heat-Induced Epitope Retrieval Using The Decloaker |
|----|---|
| | Add 500 ml of distilled water to the pan inside the decloaker. |
| | Place a full rack of slides into a Tissue Tek® container with 200 ml of 1X citrate buffer |
| | (Insert blank slides into any empty slots in the rack to ensure even heating of slides) |
| | Place the container stably inside the pan and decloak for 5 minutes. Maximum Pressure |
| | Depressurize for 10 minutes. |
| | Remove pan top and cool for 10 minutes. Temperature Before Cooling Slides |
| | Rinse the slides in 2 changes of distilled water for 3 minutes each time. |
| | |
| 5. | Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each time. |
| | |
| | Vectastain Rabbit Elite Staining Kit |

Exp. Date ______ New Kit: yes / no

6. Apply the block from the Rabbit Elite Kit. Incubate for 20 minutes at room temperature.

| DO NOT RINS | NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK. | | | |
|--------------------|--|---|-------|--|
| 7. Avidin / Biotin | Blocking Kit | | | |
| Lot # | Exp. Date | New Kit: yes / no | | |
| | lock for 15 minutes at room to | | | |
| Quick rinse in | 1X wash buffer. | - | | |
| Apply biotin b | lock for 15 minutes at room te | emperature. | | |
| | SE SLIDES WITH BUFFER I | BEFORE ADDING PRIMARY ANTIE | BODY. | |
| 8. Apply primary | antibody at a 1:1000 dilution | . Incubate for 1 hour at room temperatu | re. | |
| Lot # | Exp. Date | | | |

For negative control slides, dilute normal rabbit IgG so that it's IgG protein concentration matches that

| of the primary antibody (if necessary). Then make a 1:1000 dilution. If the concentrations can't be matched using this method, the dilution for the negative reagent may need to be adjusted. Apply the negative and incubate for 1 hour at room temperature. Lot # Exp. Date | | | |
|--|--|--|--|
| 9. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each. | | | |
| Apply the secondary antibody from Rabbit Elite Kit. Incubate for 30 minutes at room temperature. | | | |
| 11. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each. | | | |
| 12. Apply the label complex from the Rabbit Elite Kit. Incubate for 30 minutes at room temperature. (Prepare at least 30 minutes prior to use.) | | | |
| 13. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each. | | | |
| 14. Apply the DAB chromogen. Incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate) Lot # Exp. Date New Kit: yes / no | | | |
| 15. Rinse the slides in tap water 3 minutes. | | | |
| 16. Counterstain with hematoxylin for 20 seconds. | | | |
| 17. Rinse the slides in tap water until water is clear. | | | |
| 18. Gently agitate slides in 1X wash buffer until the tissues turn blue. | | | |
| 19. Dehydrate through the following solutions: | | | |
| | | | |

| Solutions | Repetitions | Time |
|--------------|-------------|-----------|
| 95% Ethanol | 1 time | 3 minutes |
| 100% Ethanol | 3 times | 3 minutes |
| Xylene | 2 times | 5 minutes |

20. Coverslip

Updated 07/01/08